

# dNEAT™ PCR Purification Kit

PURK-PCR-050

## Description

The dNEAT™ PCR Purification Kit provides a rapid and efficient method to purify DNA and remove contaminants from reaction mixtures (e.g. PCR, digestion or labelling reactions,). The dNEAT™ minispin columns contain an exclusive membrane that allows DNA adsorption in presence of chaotropic salts and removal of contaminants.

## Features

- ✓ Simple procedure
- ✓ 70-90% DNA recovery
- ✓ Suitable for DNA fragments as short as 75 bp
- ✓ DNA purified Ready to use in all molecular biology procedures

## Applications

- ✓ Removal of proteins and salts from PCR, restriction digestion, dephosphorylation, ligation or labelling reactions
- ✓ Changing of a restriction enzyme buffer
- ✓ Re-purification of genomic DNA

## Storage

The dNEAT™ PCR Purification Kit should be stored at room temperature (15–25°C) for up to 12 months without any reduction in performance.

## Kit Components

dNEAT™ minispin columns	50
Collection tubes (2 mL)	50
PB Buffer	25 ml
PE Buffer*	11,25 ml
EB Buffer	9 ml

\*Ethanol (96%-100%) [not included] must be added prior to use as indicated on the label. After ethanol has been added, mark the bottle to indicate that this step has been completed

### Quality Certifications

The dNEAT™ PCR Purification Kit is tested in the purification of a 0.6 kb DNA fragment from PCR mixture. The purified band is analysed in agarose gel electrophoresis.

### Product use limitation

This product is developed, designed and sold exclusively for research purposes and use only. The product is not intended for diagnostics or drug development, nor is it suitable for administration to humans or animals.

## Protocol

1. Add 5 volumes of PB Buffer to one volume of PCR solution and mix thoroughly by pipetting
2. Label the lid of a new spin column placed in a 2 ml collection tube. Carefully transfer the mix from step 1 to the spin column and centrifuge at 13000 rpm for 1 minute
3. Place the spin column in a new 2 ml collection tube and discard the collection tube containing the filtrate
4. Add 700  $\mu$ l of PE buffer to the minispin column and centrifuge at 13000 rpm for 1 minute  
*Remember! Before using it for the first time, add ethanol (96–100%) to the PE Buffer as indicated on the bottle*
5. Discard the flow-through and centrifuge at 13000 rpm for 1 minute  
*This step is essential for removing traces of PE buffer*
6. Place the minispin column into a new, labelled 1.5 ml microcentrifuge tube
7. Carefully open the minispin column and add 30  $\mu$ l Buffer EB or H<sub>2</sub>O (pH=7.0-8.5) directly onto the membrane. Close the cap and incubate for 1 min at room temperature, then centrifuge at 13000 rpm for 1 min to elute DNA

*To increase the DNA yield you can warm the buffer EB/H<sub>2</sub>O to 65 °C before adding it to the column*

